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Gene Section Review

IL1RN (interleukin 1 receptor antagonist)

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Published in Atlas Database: March 2014

Online updated version : http://AtlasGeneticsOncology.org/Genes/IL1RNID40953ch2q13.html DOI: 10.4267/2042/54137

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Abstract

Review on IL1RN, with data on DNA/RNA, on the protein encoded and where the gene is implicated.

Identity

Other names: DIRA, ICIL-1RA, IL-1RN, IL-1ra, IL-1ra3, IL1F3, IL1RA, IRAP, MVCD4

HGNC (Hugo): IL1RN

Location: 2q13

Local order

IL1RN is located between PSD4 gene (pleckstrin and Sec7 domain containing 4) and IL36RN (interleukin 36 receptor antagonist) and IL1F10 (interleukin 1 family, member 10 (theta)) genes (NCBI).

DNA/RNA

Note

IL1RN (interleukin 1 receptor antagonist) was identified in 1990 by Carter and cols (Carter et al., 1990).

It codes a protein that binds to interleukin 1 receptor (IL1R1) and inhibits the binding of interleukin 1 alpha and beta (IL1A and IL1B), blocking the biological activity of these two cytokines, this is the first interleukin 1 family member described that has antagonist function (Arend, 1991; Arend and Gabay, 2000).



IL1RN locus is 2q13. The gene length is 22901 bp (starts at 113868692 - ends at 113891592, according NCBI 08-Jul-2013). In this region is located along with IL36RN, IL1F10 and PSD4 and PAX8.

revues



Linear diagram of IL1RN gene with their 6 exons (pink boxes).

This gene is overexpressed in different infectious diseases and immune conditions.

The IL1RN gene has a length of 22901 base-pairs, and is constituted by 6 exons; there are four isoforms described to date that are produced by alternative splicing: one isoform is secreted and the other three are cytoplasmic (Gabay et al., 1997; Arend and Guthridge, 2000).

Description

The IL1RN gene has a length of 22901 base-pairs, encodes a member of the interleukin 1 cytokine family. The gene is located on 2q13.

Transcription

6 exons; mRNA linear (NM_173842.2) with 1794 bp. IL1RN mRNA does not contain the AUUUA sequence that has been implicated in shortening the half-life of several cytokine mRNAs (Carter et al., 1990).

Pseudogene

No pseudogenes have been identified.

Protein

Note

The IL1RN precursor protein consists of 177 amino acids and has a molecular weight of 20055 Daltons. The IL1RN cDNA encodes a 152 amino acid protein preceded by a 25 amino acid secretory leader sequence indicating that this protein takes a more straightforward pathway out of the cell than IL1.

IL1RN is found extracellularly in its mature form without requiring extracellular cleavage to its mature form. The mature protein consists in ~159 amino acids (Arend, 1991; Haskill et al., 1991).

The IL1RN gene produces two forms of IL1RN: intracellular (icIL1RN) and secreted (sIL1RN) which are controlled by different promoter regions (Redlitz et al., 2004).

Description

The protein consists of 177 amino acids, and a molecular weight of 20KDa. The amino acid sequence homology is 26-30% to IL1 β and 19% to IL1 α . The mature protein is a single nonglycosylated polypeptide of 150 amino acids approximately, further, this protein contains a 25 amino acids leader sequence (Eisenberg et al., 1990).

Expression

Practically, IL1RN is expressed in whole organism, in both adult and embryonic stages. IL1RN is expressed physiologically in different tissues like lymph node, brain, heart, colon, adipocyte, kidney, liver, lung, thyroid, adrenal gland, skin, placenta, ovary, prostate and testis (GeneCards). Furthermore, IL1RN is present in numerous cancers such as gastric cancer (Iizuka et al., 1999), cervical cancer (Fujiwaki et al., 2003), lymphoblastic (Hulkkonen et al., 2000) and myelogenous (Estrov et al., 1992) leukemias, breast cancer (Miller et al., 2000), endometrial cancer (Van Le et al., 1991), bladder cancer (Ahirwar et al., 2009), colorectal cancer (Viet et al., 2005), lung cancer (Lind et al., 2005) and brain tumors (Oelmann et al., 1997; Ilyin et al., 1998).

Localisation

There are four isoforms of IL1RN. The IL1RN isoform 1 is secreted, and the other three, IL1RN2, IL1RN3 and IL1RN4 have localization into cytoplasm (Arend and Guthridge, 2000).

IL1RN isoform	Number of Exons	Length	Protein Sequence	
			MEICRGLRSH LITLLLFLFH SETICRPSGR KSSKMQAFRI WDVNQKTFYL RNNQLVF	AGYL
1	4	1794pb	QGPNVNLEEK IDVVPIEPHA LFLGIHGGKM CLSCVKSGDE TRLQLEAVNI TDLSENF	RKQD 177aa
			KRFAFIRSDS GPTTSFESAA CPGWFLCTAM EADQPVSLTN MPDEGVMVTK FYFQEDE	2
			MALETICRPS GRKSSKMQAF RIWDVNQKTF YLRNNQLVAG YLQGPNVNLE EKIDVVE	PIEP
2	6	1865pb	HALFLGIHGG KMCLSCVKSG DETRLQLEAV NITDLSENRK QDKRFAFIRS DSGPTTS	SFES 159aa
			AACPGWFLCT AMEADQPVSL TNMPDEGVMV TKFYFQEDE	
			MALADLYEEG GGGGGEGEDN ADSKETICRP SGRKSSKMQA FRIWDVNQKT FYLRNNG	QLVA
3	5	1802pb	GYLQGPNVNL EEKIDVVPIE PHALFLGIHG GKMCLSCVKS GDETRLQLEA VNITDLS	SENR 180aa
			KQDKRFAFIR SDSGPTTSFE SAACPGWFLC TAMEADQPVS LINMPDEGVM VTKFYFQ	QEDE
			MQAFRIWDVN QKTFYLRNNQ LVAGYLQGPN VNLEEKIDVV PIEPHALFLG IHGGKMC	CLSC
4	6	1973pb	VKSGDETRLQ LEAVNITDLS ENRKQDKRFA FIRSDSGPTT SFESAACPGW FLCTAME	ADQ 143aa
			PVSLTNMPDE GVMVTKFYFQ EDE	

Table of four IL1RN isoforms, showing the amino acid sequences and other protein characteristics.



When IL1 binds to IL1-receptor induces proinflammatory reaction and gene expression. Blocking of the receptor by IL1RN prevents this response.

Function

The protein binds to interleukin 1 receptor (IL1R1) and inhibits the binding of interleukin 1 alpha and beta (IL1A and IL1B) (Arend, 1991; Arend and Gabay, 2000), modulating the immune response (Dinarello, 2011).

IL1RN competes with IL1 for binding two IL1 cell surface receptors type I and type II (IL1RtI and IL1RtII), however, when it occupies the receptor it does not trigger the cellular responses typical of IL1 which includes the production of secondary substances that mediate inflammatory responses and tissue remodeling (Dinarello, 2011).

Homology

Homologs of IL1RN protein are highly conserved in different species (NCBI). Rattus norvegicus: Il1rn (178 amino acids). Mus musculus: Il1rn (159 aa). Equus caballus: IL1RN (177 aa). Bos taurus: IL1RN (177 aa). Canis lupus familiaris: IL1RN (176 aa). Tursiops truncatus: IL1RN (177 aa). Gallus gallus: IL1RN (163 aa). Macaca fascicularis: IL1RN (177 aa).

Mutations

Note

Allelic variants

Germinal

The most common variation in the IL1RN gene is the penta allelic variable number of tandem repeat of 86 base pair located in intron 2 which results in a short allele with two repeats (IL1RN*2), and long alleles (IL1RN*L): allele 1 (four repeats), allele 3 (five repeats), allele 4 (three repeats) and allele 5 (six repeats).

The most frequent allelic form is allele 1 followed by allele 2, the rest of the alleles are very rare.

It has been reported that the allele 2 causes a 10fold increased seric IL1RN (Danis et al., 1995). Furthermore, Redlitz and cols, found that allele 1 has a 4-fold increase of the production of icIL1RN compared to allele 1 (Redlitz et al., 2004).

Two other germline mutations: Gln57Ter (rs121913162) and Glu80Ter (rs121913161) have been associated with the deficiency of IL1RN leaning to an autoinflammatory (Aksentijevich et al., 2009).

Somatic

The somatic mutation rs148026279 producing the protein change Phe148Val has been associated with malignant melanoma (Wei et al., 2011).

Implicated in

Various cancers

Note

The IL1 cluster has been strongly associated with different types of cancer. Multiple studies have shown that IL1RN, included in this cluster is altered in cancer development. IL1RN plays a central role in the response to pathogens associated with cancer etiopathogenesis (Roberge et al., 1996; Hurme and Helminen, 1998; Wang et al., 2003; Queiroz et al., 2004; Rocha et al., 2005) and chronic inflammation, well described as an important risk factor in cancer development (Hanahan and Weinberg, 2011; Baniyash et al., 2014; Khatami, 2014), thus, cytokines such as IL1RN could be used as risk and progression markers in the future.

In the meta-analysis performed by Zhang and cols., which included 71 studies of multiple cancers, with 14854 cases and 19337 controls, the research group found a consistent association with gastric cancer. IL1RN polymorphisms frequency is significantly different across ethnicities; the frequency of allele 2 is significantly lower in Asian controls (11.14%) compared to Caucasian controls (26%) (Zhang et al., 2011).

Gastric cancer

Note

Gastric cancer after H. pylori infection susceptibility has been linked with IL1RN gene. Studies of genetic association have shown an increased risk of gastric cancer with the allele IL1RN*2 across different populations (El-Omar et al., 2000; Furuta et al., 2002; Alpizar-Alpizar et al., 2005; Garza-Gonzalez et al., 2005; Palli et al., 2005; Morgan et al., 2006; Oliveira et al., 2012). However, IL1RN*2 has been also been associated independently of H. pylori infection (Mattar et al., 2013).

Oncogenesis

The inflammatory response after H. pylori infection has been linked to the IL1 gene cluster. The infection induces the synthesis of IL1B which binds to its target receptor and starts the inflammatory response against the pathogen (Sierra et al., 2008). The IL1RN competitively binds to the IL1 receptor with the same affinity as IL1 without activating the inflammation cascade, modulating the effects of IL1B. The severity of damage caused by inflammatory response in mucosal tissue is regulated by the balance between these cytokines. The short allele IL1RN*2, is associated with increased levels of IL1B, which could probably have a significant effect on increased inflammatory damage (Santtila et al., 1998).

It has also been described that IL1B inhibits gastric secretion (Sugimoto et al., 2009), and it is 100 times more potent than proton pump inhibitor (Wolfe and Nompleggi, 1992), which favors the corpus colonization, causing atrophic gastritis that may evolve to gastric carcinoma. El-Omar and cols., demonstrated an increased risk of hypochlorhydria when IL1RN*2 allele is present in homozygous form (El-Omar et al., 2000).

Cervical cancer

Oncogenesis

Cervical cancer is associated with the infection by human papillomavirus (HPV), however, although millions of women are infected with high-risk HPV subtypes, only a subset of them develop cervical cancer, reveling an important role of host immunity in cervical carcinogenesis. Various studies have observed a significant contribution of IL1RN*2 allele to increase risk of cervical cancer (Sehouli et al., 2002; Mustea et al., 2003; Tamandani et al., 2008; Sousa et al., 2012). Moreover, Sousa and cols, found that IL1RN*2 correlated not only with cervical cancer but with cervical lesions and earlier onset of cases with cervical lesion and cancer in patients homozygous IL1RN*2 (Sousa et al., 2012). Tamandani and cols, showed a protective association of heterozygous IL1RN*1*2 and homozygous IL1RN*2 and HPV 16 and 18 subtypes but а risk association with adenocarcinoma (Tamandani et al., 2008).

Breast cancer

Prognosis

In Caucasian women it has been described that IL1RN*2 is associated with shortened disease free survival and overall survival (Grimm et al., 2009). In this study, Grimm and cols reported that only 80% of women positive to IL1RN*2 survived after 12 months, and 30% had died after 48 months; the disease-free survival was only of 40% in women with IL1RN*2 compared with 80% in women with the wild allele (Grimm et al., 2009). In respect to the long alleles of IL1RN gene, allele *2 has been reported to be modify the binding on the IL1 receptor, leading to less efficient inhibition of IL1a and IL1b (Tarlow et al., 1993), this might result in a proinflammatory status and enhanced tumor aggressiveness, which is likely to result in a shortened survival of women with breast cancer (Grimm et al., 2009).

Oncogenesis

Breast cancer oncogenesis has been associated with polymorphisms in different cytokines (Gomez-Flores-Ramos et al., 2013; Dinarello, 2006) and it has been widely discuss the participation of chronic inflammation in breast carcinogenesis (Honma et al., 2002). Different research groups have studied the association of IL1RN and breast cancer risk. Lee and cols, found a decreased breast cancer risk with the short allele (*2) and a higher risk of cancer in women with the long allele and higher body mass index (Lee et al., 2006). Zhang and cols, performed a meta-analysis study and found a similar trend with allele IL1RN*2 (Zhang et al., 2011), however, with this apparent diminish in risk for breast cancer, other studies have found an elevated risk of earlier recurrence of breast cancer in women with IL1RN*2 allele (Grimm et al., 2009).

Bladder cancer

Oncogenesis

Significant association with higher risk of bladder cancer has been described to II1RN*2 allele (Bid et al., 2006; Ahirwar et al., 2009). This allele has been proposed as a potential marker for genetic susceptibility to bladder cancer (Ahirwar et al., 2009). Studies have showed that IL1RN*2 increases the production of IL1B significantly (Santtila et al., 1998; Nazarenko et al., 2008) and it can induce angiogenesis via upregulation of COX-2 or inducible nitric oxide and vascular endothelial growth factor which may contribute to tumor growth (Rahman et al., 2001).

Colorectal cancer

Prognosis

IL1B and IL1RN have been shown to play an important role in angiogenesis of early onset tumors, Viet and cols, found that allele *1 was more frequent in patients with localized disease compared with disseminated disease, being allele *2 associated with dissemination of the disease (Viet et al., 2005). The study of Lurje and cols, showed that patients with IL1RN VNTR had a significant six-fold increment in relative risk of developing tumor recurrence compared to those patients with the wild allele. The IL1RN homozygous *2/*2 genotype had a median time-to-recurrence of 5.7 years, compared with 10.7 years for those with *1/*1 genotype (Lurje et al., 2009).

Serum levels of IL1Ra have been studies as prognostic factors as well, in colorectal cancer patients, low preoperative IL1Ra was associated with postoperative infection (Miki et al., 2005).

Oncogenesis

Colorectal cancer has been widely associated in epidemiological and experimental studies with chronic inflammation. Viet and cols, found that the allele IL1RN*3 of VNTR variant was significantly increased in patients compared with controls, and the allelic distribution of this VNTR differed between colon and rectum, being allele *3 more abundant in colon (Viet et al., 2005). Other polymorphic sites have been associated with colorectal oncogenesis. Burada and cols, linked the polymorphism IL1RN +2018C>T with colorectal cancer, allele C was found to be enriched in patients with cancer compared to controls. This study described that this association was limited to early stage I and II (Burada et al., 2013).

In attempt to find markers for colorectal cancer disease, serum cytokines levels have been studied; Iwagaki and cols, found that patients with colorectal cancer present reduced level of IL1Ra relative to normal controls, indicating that cancer patients have an immunologic disorder (Iwagaki et al., 1997).

Lung cancer

Oncogenesis

Tobacco smoking is the main risk factor for lung cancer, however, only 10-15% of smokers develop lung cancer, suggesting that genetic factors are important in individual susceptibility for this disease (Ridge et al., 2013). Lind and cols, found that individuals homozygous for IL1RN*1 in combination with the allele IL1B-31T had an increased risk of non-small cell lung cancer and a two-fold higher level of bulky/hydrophobic DNA adducts in the long in patients with IL1RN*1 (Lind et al., 2005). These data were confirmed in Chinese patients, with a decreased risk of 32% in patients with IL1RN*2 allele (Hu et al., 2006). On the other hand, Lim and cols, found a 5-fold-time increased risk in lung cancer in never-smokers patients with the IL1RN*2 (Lim et al., 2011), but these results are not consistent, since Hu and cols, found a reduced risk in non-smokers with squamous cell carcinoma (Hu et al., 2006).

Brain cancer

Oncogenesis

Inflammatory status of brain tumors has been studied, and it has been shown that IL1 cluster is important in development and progression of neoplasia. Ilyin and cols, investigated the levels of IL1 and IL1Ra in pediatric astrocytomas, ependymomas and primitive neuroectodermal tumors. The results demonstrated a significant different profile among tumors. Pilocytic, nonpilocytic and anaplastic astrocytomas had a significant increase of mRNA of IL1 beta and its receptor, but low levels of IL1Ra mRNA, suggesting an imbalance between stimulatory and inhibitory cytokines in brain tumors growth and development via autocrine/paracrine mechanisms (Ilyin et al., 1998).

Oelmann and cols, performed a study in cell lines of glioblastoma showed that IL1Ra modulates glioblastoma growth. Experimental addition of neutralizing antibody against IL1Ra downregulated growth of IL1 and IL1Ra producing glioblastoma, the authors suggest that an autocrine production of IL1Ra can counteract IL1 function and represent a basic escape mechanism malignant growth in some glioblastomas (Oelmann et al., 1997).

Septic shock in pediatric population with acute lymphoblastic leukemia

Prognosis

The presence of IL1RN*2 allele was associated with significant susceptibility to septic shock in pediatric patients with acute lymphoblastic leukemia by (Zapata-Tarres et al., 2013). The patients studied by Zapata-Tarres and cols, were susceptible to septic shock.

The association between sepsis and IL1RN*2 has been reported previously, Fang and cols, reported an increased relative risk of sepsis in patients homozygous IL1RN*2 as well heterozygous patients (Fang et al., 1999). Arnalich and cols reported a significant increase in the risk of death after severe sepsis in patients with IL1RN*2 and that the allele is associated with decreased production of IL1Ra in culture but higher concentrations of the protein in serum (Arnalich et al., 2002).

References

Carter DB, Deibel MR Jr, Dunn CJ, Tomich CS, Laborde AL, Slightom JL, Berger AE, Bienkowski MJ, Sun FF, McEwan RN. Purification, cloning, expression and biological characterization of an interleukin-1 receptor antagonist protein. Nature. 1990 Apr 12;344(6267):633-8

Eisenberg SP, Evans RJ, Arend WP, Verderber E, Brewer MT, Hannum CH, Thompson RC. Primary structure and functional expression from complementary DNA of a human interleukin-1 receptor antagonist. Nature. 1990 Jan 25;343(6256):341-6

Arend WP. Interleukin 1 receptor antagonist. A new member of the interleukin 1 family. J Clin Invest. 1991 Nov;88(5):1445-51

Haskill S, Martin G, Van Le L, Morris J, Peace A, Bigler CF, Jaffe GJ, Hammerberg C, Sporn SA, Fong S. cDNA cloning of an intracellular form of the human interleukin 1 receptor antagonist associated with epithelium. Proc Natl Acad Sci U S A. 1991 May 1;88(9):3681-5

Van Le L, Haskill S, Jaffe GJ, Fowler WC Jr. Expression of interleukin-1 and interleukin-1 receptor antagonists in endometrial cancer. Gynecol Oncol. 1991 Aug;42(2):161-4

Estrov Z, Kurzrock R, Estey E, Wetzler M, Ferrajoli A, Harris D, Blake M, Gutterman JU, Talpaz M. Inhibition of acute myelogenous leukemia blast proliferation by interleukin-1 (IL-1) receptor antagonist and soluble IL-1 receptors. Blood. 1992 Apr 15;79(8):1938-45

Wolfe MM, Nompleggi DJ. Cytokine inhibition of gastric acid secretion--a little goes a long way. Gastroenterology. 1992 Jun;102(6):2177-8

Tarlow JK, Blakemore AI, Lennard A, Solari R, Hughes HN, Steinkasserer A, Duff GW. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by

variable numbers of an 86-bp tandem repeat. Hum Genet. 1993 May;91(4):403-4

Danis VA, Millington M, Hyland VJ, Grennan D. Cytokine production by normal human monocytes: inter-subject variation and relationship to an IL-1 receptor antagonist (IL-1Ra) gene polymorphism. Clin Exp Immunol. 1995 Feb;99(2):303-10

Roberge CJ, Poubelle PE, Beaulieu AD, Heitz D, Gosselin J. The IL-1 and IL-1 receptor antagonist (IL-1Ra) response of human neutrophils to EBV stimulation. Preponderance of IL-Ra detection. J Immunol. 1996 Jun 15;156(12):4884-91

Gabay C, Smith MF, Eidlen D, Arend WP. Interleukin 1 receptor antagonist (IL-1Ra) is an acute-phase protein. J Clin Invest. 1997 Jun 15;99(12):2930-40

Iwagaki H, Hizuta A, Tanaka N. Interleukin-1 receptor antagonists and other markers in colorectal cancer patients. Scand J Gastroenterol. 1997 Jun;32(6):577-81

Oelmann E, Kraemer A, Serve H, Reufi B, Oberberg D, Patt S, Herbst H, Stein H, Thiel E, Berdel WE. Autocrine interleukin-1 receptor antagonist can support malignant growth of glioblastoma by blocking growth-inhibiting autocrine loop of interleukin-1. Int J Cancer. 1997 Jun 11;71(6):1066-76

Hurme M, Helminen M. Polymorphism of the IL-1 gene complex in Epstein-Barr virus seronegative and seropositive adult blood donors. Scand J Immunol. 1998 Sep;48(3):219-22

Ilyin SE, González-Gómez I, Gilles FH, Plata-Salamán CR. Interleukin-1 alpha (IL-1 alpha), IL-1 beta, IL-1 receptor type I, IL-1 receptor antagonist, and TGF-beta 1 mRNAs in pediatric astrocytomas, ependymomas, and primitive neuroectodermal tumors. Mol Chem Neuropathol. 1998 Feb;33(2):125-37

Santtila S, Savinainen K, Hurme M. Presence of the IL-1RA allele 2 (IL1RN*2) is associated with enhanced IL-1beta production in vitro. Scand J Immunol. 1998 Mar;47(3):195-8

Fang XM, Schröder S, Hoeft A, Stüber F. Comparison of two polymorphisms of the interleukin-1 gene family: interleukin-1 receptor antagonist polymorphism contributes to susceptibility to severe sepsis. Crit Care Med. 1999 Jul;27(7):1330-4

lizuka N, Hazama S, Hirose K, Abe T, Tokuda N, Fukumoto T, Tangoku A, Oka M. Interleukin-1 receptor antagonist mRNA expression and the progression of gastric carcinoma. Cancer Lett. 1999 Aug 3;142(2):179-84

Arend WP, Gabay C. Physiologic role of interleukin-1 receptor antagonist. Arthritis Res. 2000;2(4):245-8

Arend WP, Guthridge CJ. Biological role of interleukin 1 receptor antagonist isoforms. Ann Rheum Dis. 2000 Nov;59 Suppl 1:i60-4

EI-Omar EM, Carrington M, Chow WH, McColl KE, Bream JH, Young HA, Herrera J, Lissowska J, Yuan CC, Rothman N, Lanyon G, Martin M, Fraumeni JF Jr, Rabkin CS. Interleukin-1 polymorphisms associated with increased risk of gastric cancer. Nature. 2000 Mar 23;404(6776):398-402

Hulkkonen J, Vilpo J, Vilpo L, Koski T, Hurme M. Interleukin-1 beta, interleukin-1 receptor antagonist and interleukin-6 plasma levels and cytokine gene polymorphisms in chronic lymphocytic leukemia: correlation with prognostic parameters. Haematologica. 2000 Jun;85(6):600-6 Miller LJ, Kurtzman SH, Anderson K, Wang Y, Stankus M, Renna M, Lindquist R, Barrows G, Kreutzer DL. Interleukin-1 family expression in human breast cancer: interleukin-1 receptor antagonist. Cancer Invest. 2000;18(4):293-302

Rahman MA, Dhar DK, Yamaguchi E, Maruyama S, Sato T, Hayashi H, Ono T, Yamanoi A, Kohno H, Nagasue N. Coexpression of inducible nitric oxide synthase and COX-2 in hepatocellular carcinoma and surrounding liver: possible involvement of COX-2 in the angiogenesis of hepatitis C virus-positive cases. Clin Cancer Res. 2001 May;7(5):1325-32

Arnalich F, López-Maderuelo D, Codoceo R, Lopez J, Solis-Garrido LM, Capiscol C, Fernandez-Capitán C, Madero R, Montiel C. Interleukin-1 receptor antagonist gene polymorphism and mortality in patients with severe sepsis. Clin Exp Immunol. 2002 Feb;127(2):331-6

Furuta T, El-Omar EM, Xiao F, Shirai N, Takashima M, Sugimura H. Interleukin 1beta polymorphisms increase risk of hypochlorhydria and atrophic gastritis and reduce risk of duodenal ulcer recurrence in Japan. Gastroenterology. 2002 Jul;123(1):92-105

Honma S, Shimodaira K, Shimizu Y, Tsuchiya N, Saito H, Yanaihara T, Okai T. The influence of inflammatory cytokines on estrogen production and cell proliferation in human breast cancer cells. Endocr J. 2002 Jun;49(3):371-7

Sehouli J, Mustea A, Könsgen D, Katsares I, Lichtenegger W. Polymorphism of IL-1 receptor antagonist gene: role in cancer. Anticancer Res. 2002 Nov-Dec;22(6A):3421-4

Fujiwaki R, lida K, Nakayama K, Kanasaki H, Hata K, Katabuchi H, Okamura H, Miyazaki K. Clinical significance of interleukin-1 receptor antagonist in patients with cervical carcinoma. Gynecol Oncol. 2003 Apr;89(1):77-83

Mustea A, Sehouli J, Könsgen D, Stengel D, Sofroni D, Lichtenegger W. Interleukin 1 receptor antagonist (IL-1RA) polymorphism in women with cervical cancer. Anticancer Res. 2003 Mar-Apr;23(2A):1099-102

Wang Y, Kato N, Hoshida Y, Yoshida H, Taniguchi H, Goto T, Moriyama M, Otsuka M, Shiina S, Shiratori Y, Ito Y, Omata M. Interleukin-1beta gene polymorphisms associated with hepatocellular carcinoma in hepatitis C virus infection. Hepatology. 2003 Jan;37(1):65-71

Queiroz DM, Guerra JB, Rocha GA, Rocha AM, Santos A, De Oliveira AG, Cabral MM, Nogueira AM, De Oliveira CA. IL1B and IL1RN polymorphic genes and Helicobacter pylori cagA strains decrease the risk of reflux esophagitis. Gastroenterology. 2004 Jul;127(1):73-9

Redlitz KH, Yamshchikov VF, Cominelli F. Differential contribution of IL-1Ra isoforms to allele-specific IL-1Ra mRNA accumulation. J Interferon Cytokine Res. 2004 Apr;24(4):253-60

Alpízar-Alpízar W, Pérez-Pérez GI, Une C, Cuenca P, Sierra R. Association of interleukin-1B and interleukin-1RN polymorphisms with gastric cancer in a high-risk population of Costa Rica. Clin Exp Med. 2005 Dec;5(4):169-76

Garza-González E, Bosques-Padilla FJ, El-Omar E, Hold G, Tijerina-Menchaca R, Maldonado-Garza HJ, Pérez-Pérez GI. Role of the polymorphic IL-1B, IL-1RN and TNF-A genes in distal gastric cancer in Mexico. Int J Cancer. 2005 Mar 20;114(2):237-41

Lind H, Zienolddiny S, Ryberg D, Skaug V, Phillips DH, Haugen A. Interleukin 1 receptor antagonist gene polymorphism and risk of lung cancer: a possible interaction with polymorphisms in the interleukin 1 beta gene. Lung Cancer. 2005 Dec;50(3):285-90

Miki C, Inoue Y, Toiyama Y, Ojima E, Kobayashi M, Hatada T, Araki T, Kusunoki M. Deficiency in systemic interleukin-1 receptor antagonist production as an operative risk factor in malnourished elderly patients with colorectal carcinoma. Crit Care Med. 2005 Jan;33(1):177-80

Palli D, Saieva C, Luzzi I, Masala G, Topa S, Sera F, Gemma S, Zanna I, D'Errico M, Zini E, Guidotti S, Valeri A, Fabbrucci P, Moretti R, Testai E, del Giudice G, Ottini L, Matullo G, Dogliotti E, Gomez-Miguel MJ. Interleukin-1 gene polymorphisms and gastric cancer risk in a high-risk Italian population. Am J Gastroenterol. 2005 Sep;100(9):1941-8

Rocha GA, Guerra JB, Rocha AM, Saraiva IE, da Silva DA, de Oliveira CA, Queiroz DM. IL1RN polymorphic gene and cagA-positive status independently increase the risk of noncardia gastric carcinoma. Int J Cancer. 2005 Jul 10;115(5):678-83

Viet HT, Wågsäter D, Hugander A, Dimberg J. Interleukin-1 receptor antagonist gene polymorphism in human colorectal cancer. Oncol Rep. 2005 Oct;14(4):915-8

Bid HK, Manchanda PK, Mittal RD. Association of interleukin-1Ra gene polymorphism in patients with bladder cancer: case control study from North India. Urology. 2006 May;67(5):1099-104

Dinarello CA. The paradox of pro-inflammatory cytokines in cancer. Cancer Metastasis Rev. 2006 Sep;25(3):307-13

Hu Z, Shao M, Chen Y, Zhou J, Qian J, Xu L, Ma H, Wang X, Xu Y, Lu D, Shen H. Allele 2 of the interleukin-1 receptor antagonist gene (IL1RN*2) is associated with a decreased risk of primary lung cancer. Cancer Lett. 2006 May 18;236(2):269-75

Lee KM, Park SK, Hamajima N, Tajima K, Choi JY, Noh DY, Ahn SH, Yoo KY, Hirvonen A, Kang D. Genetic polymorphisms of interleukin-1 beta (IL-1B) and IL-1 receptor antagonist (IL-1RN) and breast cancer risk in Korean women. Breast Cancer Res Treat. 2006 Mar;96(2):197-202

Morgan DR, Dominguez RL, Keku TO, Heidt PE, Martin CF, Galanko JA, Omofoye OA, Sandler RS. Gastric cancer and the high combination prevalence of host cytokine genotypes and Helicobacter pylori in Honduras. Clin Gastroenterol Hepatol. 2006 Sep;4(9):1103-11

Nazarenko I, Marhaba R, Reich E, Voronov E, Vitacolonna M, Hildebrand D, Elter E, Rajasagi M, Apte RN, Zöller M. Tumorigenicity of IL-1alpha- and IL-1beta-deficient fibrosarcoma cells. Neoplasia. 2008 Jun;10(6):549-62

Sierra R, Une C, Ramirez V, Alpizar-Alpizar W, Gonzalez MI, Ramirez JA, De Mascarel A, Cuenca P, Perez-Perez G, Megraud F. Relation of atrophic gastritis with Helicobacter pylori-CagA(+) and interleukin-1 gene polymorphisms. World J Gastroenterol. 2008 Nov 14;14(42):6481-7

Tamandani DM, Sobti RC, Shekari M, Kaur S, Huria A. Impact of polymorphism in IL-1RA gene on the risk of cervical cancer. Arch Gynecol Obstet. 2008 Jun;277(6):527-33

Ahirwar DK, Agrahari A, Mandhani A, Mittal RD. Cytokine gene polymorphisms are associated with risk of urinary bladder cancer and recurrence after BCG immunotherapy.

Biomarkers. 2009 Jun;14(4):213-8

Aksentijevich I, Masters SL, Ferguson PJ, Dancey P, Frenkel J, van Royen-Kerkhoff A, Laxer R, Tedgård U, Cowen EW, Pham TH, Booty M, Estes JD, Sandler NG, Plass N, Stone DL, Turner ML, Hill S, Butman JA, Schneider R, Babyn P, El-Shanti HI, Pope E, Barron K, Bing X, Laurence A, Lee CC, Chapelle D, Clarke GI, Ohson K, Nicholson M, Gadina M, Yang B, Korman BD, Gregersen PK, van Hagen PM, Hak AE, Huizing M, Rahman P, Douek DC, Remmers EF, Kastner DL, Goldbach-Mansky R. An autoinflammatory disease with deficiency of the interleukin-1-receptor antagonist. N Engl J Med. 2009 Jun 4;360(23):2426-37

Grimm C, Kantelhardt E, Heinze G, Polterauer S, Zeillinger R, Kölbl H, Reinthaller A, Hefler L. The prognostic value of four interleukin-1 gene polymorphisms in Caucasian women with breast cancer: a multicenter study. BMC Cancer. 2009 Mar 6;9:78

Lurje G, Hendifar AE, Schultheis AM, Pohl A, Husain H, Yang D, Manegold PC, Ning Y, Zhang W, Lenz HJ. Polymorphisms in interleukin 1 beta and interleukin 1 receptor antagonist associated with tumor recurrence in stage II colon cancer. Pharmacogenet Genomics. 2009 Feb;19(2):95-102

Sugimoto M, Furuta T, Yamaoka Y. Influence of inflammatory cytokine polymorphisms on eradication rates of Helicobacter pylori. J Gastroenterol Hepatol. 2009 Nov;24(11):1725-32

Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. Blood. 2011 Apr 7;117(14):3720-32

Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011 Mar 4;144(5):646-74

Lim WY, Chen Y, Ali SM, Chuah KL, Eng P, Leong SS, Lim E, Lim TK, Ng AW, Poh WT, Tee A, Teh M, Salim A, Seow A. Polymorphisms in inflammatory pathway genes, host factors and lung cancer risk in Chinese female neversmokers. Carcinogenesis. 2011 Apr;32(4):522-9

Wei X, Walia V, Lin JC, Teer JK, Prickett TD, Gartner J, Davis S, Stemke-Hale K, Davies MA, Gershenwald JE, Robinson W, Robinson S, Rosenberg SA, Samuels Y. Exome sequencing identifies GRIN2A as frequently mutated in melanoma. Nat Genet. 2011 May;43(5):442-6

Zhang B, Beeghly-Fadiel A, Long J, Zheng W. Genetic variants associated with breast-cancer risk: comprehensive research synopsis, meta-analysis, and epidemiological evidence. Lancet Oncol. 2011 May;12(5):477-88

Oliveira JG, Duarte MC, Silva AE. IL-1ra anti-inflammatory cytokine polymorphism is associated with risk of gastric cancer and chronic gastritis in a Brazilian population, but the TNF- β pro-inflammatory cytokine is not. Mol Biol Rep. 2012 Jul;39(7):7617-25

Sousa H, Santos AM, Catarino R, Pinto D, Moutinho J, Canedo P, Machado JC, Medeiros R. IL-1RN VNTR polymorphism and genetic susceptibility to cervical cancer in Portugal. Mol Biol Rep. 2012 Dec;39(12):10837-42

Burada F, Dumitrescu T, Nicoli R, Ciurea ME, Angelescu C, Mixich F, Ioana M. IL-1RN +2018T>C polymorphism is correlated with colorectal cancer. Mol Biol Rep. 2013 Apr;40(4):2851-7

Gómez Flores-Ramos L, Escoto-De Dios A, Puebla-Pérez AM, Figuera-Villanueva LE, Ramos-Silva A, Ramírez-Patiño R, Delgado-Saucedo JI, Salas-González E, Zúñiga-González GM, Alonzo-Rojo A, Gutiérrez-Hurtado I, Gallegos-Arreola MP. Association of the tumor necrosis factor-alpha -308G>A polymorphism with breast cancer in Mexican women. Genet Mol Res. 2013 Nov 18;12(4):5680-93

Mattar R, Marques SB, Dos Santos AF, do Socorro Monteiro M, Iriya K, Carrilho FJ. A possible role of IL-1RN gene polymorphism in the outcome of gastrointestinal diseases associated with H. pylori infection. Clin Exp Gastroenterol. 2013;6:35-41

Ridge CA, McErlean AM, Ginsberg MS. Epidemiology of lung cancer. Semin Intervent Radiol. 2013 Jun;30(2):93-8

Zapata-Tarrés M, Arredondo-García JL, Rivera-Luna R, Klünder-Klünder M, Mancilla-Ramírez J, Sánchez-Urbina R, Vázquez-Cruz MY, Juárez-Villegas LE, Palomo-Colli MA. Interleukin-1 receptor antagonist gene polymorphism increases susceptibility to septic shock in children with acute lymphoblastic leukemia. Pediatr Infect Dis J. 2013 Feb;32(2):136-9

Baniyash M, Sade-Feldman M, Kanterman J. Chronic inflammation and cancer: suppressing the suppressors. Cancer Immunol Immunother. 2014 Jan:63(1):11-20

Khatami M. Chronic Inflammation: Synergistic Interactions of Recruiting Macrophages (TAMs) and Eosinophils (Eos) with Host Mast Cells (MCs) and Tumorigenesis in CALTs. M-CSF, Suitable Biomarker for Cancer Diagnosis! Cancers (Basel). 2014 Jan 27;6(1):297-322

This article should be referenced as such:

Gómez-Flores-Ramos L, Torres-Flores J, Gallegos-Arreola MP. IL1RN (interleukin 1 receptor antagonist). Atlas Genet Cytogenet Oncol Haematol. 2014; 18(10):746-753.