AAMP (angio-associated, migratory cell protein)

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Identity
HGNC (Hugo): AAMP
Location: 2q35

DNA/RNA
Note
The NCBI RefSeq (Aug-2011) consensus sequence for AAMP differs at the 5-prime end from the manually sequenced version published initially. A diagram of the NCBI RefSeq (Aug-2011) version for rAAMP is shown here.

Description
The AAMP gene (NCBI RefSeq, Aug-2011) encompasses 6042 bp; 11 exons.

Transcription
1859 bp mRNA (NCBI RefSeq, Aug-2011).

Pseudogene
None are known.

rAAMP encoding a 434 aa protein in normal cells. The codon for AAMP's initiating methionine, the stop codon, and the polyadenylation sites are indicated at 85-87, 1387-1389, and 1774-1779, respectively. Untranslated sequence is indicated in the hatched regions at the 3' end of exon 1 and the 5' end of exon 11. The poly-adenosine tail, 1793-1859, is included.

The bulk of AAMP's sequence is constituted by WD40 domains that are known to commonly fold to form a platform for active portions of a protein. Thus the stretch of contiguous glutamic acid residues should be available for interactions with other proteins. Also, binding of other proteins may occur to regions of these repeats. The WD40 repeats are known to mediate aggregation of subunits to form complex, multi-protein structures. Two immunoglobulin-type domains are designated by pairs of cysteines, (C96 - C130) and (C216 - C265). AAMP may play a negative role in Nod2-mediated NF-κB activation via a physical interaction in the region indicated. A physical interaction between AAMP and thromboxane A2 receptors (TPalpha and TPhbeta) has been suggested to occur via multiple sites with no particular domain identified in localization studies.
**Protein**

**Note**
The NCBI RefSeq (Aug-2011) consensus sequence for AAMP (434 aa) is shorter than initially reported (452 aa, 52 kDa).

**Description**
434 amino acids, 49 kDa protein (NCBI RefSeq, Aug-2011).

**Expression**
AAMP is widely expressed among many types of mammalian cells and has been conserved in evolution.

**Localisation**
AAMP was initially detected in the cytoplasm of many types of nucleated mammalian cells and was strongly expressed in endothelial cells, cytrophoblasts, and poorly differentiated colon adenocarcinoma cells in lymphatics. AAMP has been observed at the luminal edges of endometrial cells and has been found in the extracellular environment of vascular-associated mesenchymal cells.

**Function**
Functional studies and associations suggest roles for AAMP in angiogenesis, including endothelial tube formation, migration of endothelial and smooth muscle cells, neo-intima formation, and thromboxane A receptor interactions. In immune cells AAMP may be involved in the regulation of NF-kappaB activation mediated by Nod2. The common occurrence of WD40 domains in signaling proteins supports a signaling function as a possibility for AAMP. The epitope, ESESES, that AAMP shares with alpha-actinin and a smaller protein specific for fast skeletal muscle, suggests that it may have cytoskeletal interactions. Although AAMP was described as having heparin-binding capacity in melanoma cells, the NCBI RefSeq (Aug-2011) version of AAMP does not include the heparin-binding sequence (encodes RRLRR) in the coding sequence. The RefSeq version of AAMP identifies the initiating methionine (85-87) as being 3' to the sequence encoding RRLRR (70-84).

**Homology**
AAMP shares homology with the other members of the WD40 repeat superfamily. Several of the WD40 repeat proteins also contain an amino terminal run of glutamic acid residues outside of their WD repeats. The two immunoglobulin-type domains in AAMP resemble those of the immunoglobulin superfamily members, including domains of NCAM, DCC, NgCAM, etc.

**Mutations**

**Note**
AAMP was initially cloned and sequenced as a transcript for a 452 aa, 52 kDa protein. It was obtained from an expression library derived from melanoma cells of a brain metastasis. However the amino terminus was missing from the library clone. AAMP's amino terminus was determined by 5' RACE from human brain mRNA. Two versions were obtained and both differed from the NCBI RefSeq (Aug-2011) version of AAMP. The one shown contains coding sequence upstream from the initiating methionine in the RefSeq version. This alternative form may represent a fusion protein due to an in-frame insertion or a chromosomal rearrangement. AAMP potentially gains heparin binding functionality when sequence upstream from AUG at nucleotides 85-87 is preceded by an alternative initiating methionine codon that enables translation of the codons for RRLRR.

**Germinal**
None are known.

**Somatic**
Chromosomal rearrangements or insertions that place an initiating methionine codon further upstream than the methionine at 85-87 permit AAMP to gain heparin binding function when nucleotides 70-84 are translated.
Implicated in

**Gastrointestinal stromal tumor (GIST)**

*Note*
GISTs are the most common mesenchymal tumors of the digestive tract and are believed to arise from the interstitial cells of Cajal. They respond to imatinib, a tyrosine kinase inhibitor, which is also used to treat myeloid leukemia. Expression of AAMP was found to be increased in GISTs with mutated KIT. Expression of AAMP among various soft tissue sarcomas and normal tissues was highest in the GISTs. AAMP ranks high among the upregulated genes in GISTs.

**Myeloid leukemia (chronic (CML) and acute (AML) forms)**

*Note*
AAMP is expressed in myeloid leukemia cell lines and its expression is repressed by imatinib (mainline treatment drug for chronic myeloid leukemia), deferasirox (iron chelator that decreases cell proliferation), and anisomycin, an inducer of apoptosis.

**Lymphoma (B and T cell origins, non-Hodgkins and Hodgkins types)**

*Note*
AAMP is expressed in activated T lymphocytes, monocytes, and lymphoma cells. Compared to normal B cells, AAMP is increased in non-Hodgkins lymphomas and in classical Hodgkins lymphoma.

**Melanoma (melanocyte origin, usually skin)**

*Note*
In lysates of a melanoma cell line obtained from a brain metastasis, the size of the protein that reacted with anti-AAMP was 52 kDa, thus corresponding to the size predicted by an alternative transcript. The alternative version of AAMP could have resulted from a chromosomal rearrangement due to 2q35 breakage in the amino terminal region of AAMP. Placement of AUG at the 5' end of nucleotide 34 to serve as an alternative initiating methionine codon permits expression of the heparin binding site, RRLRR (nucleotides 70-84 in the NCBI ReqSeq (Aug-2011) version of AAMP) as a gain of function.

**Breast cancer (ductal cell origin most common)**

*Note*
Expression profiling of human breast cancer cells versus mammary epithelial cells revealed higher expression of AAMP in the tumor cells. AAMP expression is higher in ductal carcinoma in situ (DCIS) with necrosis compared to DCIS without necrosis. However, expression profiling of DCIS and invasive ductal carcinoma (IDC) paired specimens from the same patients revealed decreased AAMP in IDC. AAMP expression may be relevant for the development of DCIS.

**Glial brain tumors (neuroectodermal cell origin)**

*Note*
When compared with normal, glioblastomas (Grade IV astrocytomas) and oligodendrogliomas (Grades II - III) demonstrated slightly elevated levels of AAMP expression. Expression of AAMP was increased in drug resistant glioblastomas, primary and recurrent.

**Colon neoplasia (benign adenomas and carcinoma arise from glandular cells of the mucosa in the gastrointestinal tract)**

*Note*
Expression profiling of normal mucosa and colorectal adenomas from the same patients showed that AAMP was slightly increased in the adenomas. However, a small dataset of colon tubular adenomas harboring focal adenocarcinomas, with microdissections of paired samples from each, showed that AAMP may become decreased in the incipient carcinomas. Although AAMP may play a role in the development of colonic neoplasia, it has not been shown to be positively involved in progression of adenomas to malignancy.

**Epidermoid carcinoma (keratinocyte origin if arises in skin)**

*Note*
AAMP is expressed in squamous carcinoma cell lines. In one cell line studied with PTEN knocked down, the expression of AAMP also fell significantly. Also, for tumors from a radiosensitive cell line, AAMP expression fell significantly in radioresistant tumors derived from the sensitive cell line.

**Cervical cancer (usually arises from squamous type cells)**

*Note*
In a study of cervical carcinoma HeLa cells treated with epidermal growth factor in a time course, expression of AAMP was increased at 2-8 hours.

**Ovarian cancer (often arises from serous cells)**

*Note*
Cancer cells prepared from primary cultures of ovarian papillary serous adenocarcinomas revealed increased AAMP expression in 2 of 3 carboplatin resistant tumors whereas none of the tumors from 3 patients with sensitivity to carboplatin demonstrated comparable levels.
Papillary thyroid carcinoma (arises from cells in thyroid follicles)

Note
Analysis of papillary thyroid carcinoma tumors matched with normal thyroid from nine patients found that AAMP expression fell slightly in the tumors.

Pulmonary carcinoma (multiple types of cells can be the origin)

Note
AAMP’s expression in a lung alveolar adenocarcinoma cell line was down-regulated by TGF-beta that was added to induce an epithelial-mesenchymal transition.

Chromosomal rearrangements at 2q35

Note
The location of AAMP at 2q35 imparts susceptibility to chromosomal rearrangements involving the terminal region of chromosome 2’s long arm, q. Terminal regions are more susceptible to rearrangements than the midregions of chromosomes. Fusion transcripts and proteins result from breakage and rearrangements that occur in unstable genomes. Several fusion proteins resulting from rearrangements involving other genes in this region have been reported in association with malignancy. Rearrangements that place in-frame coding sequence upstream from nucleotides 70-84 in the NCBI RefSeq (Aug-2011) version of AAMP permit a gain of function, i.e. heparin-binding, to occur.

Breakpoints

Note
Clones obtained in 5’ RACE studies of AAMP’s amino terminus revealed a breakpoint between nucleotides 30 and 31 with inclusion of upstream sequence that did not match the NCBI RefSeq (Aug-2011) form of AAMP and also another version with a breakpoint between nucleotides 33 and 34 and introduction of an alternative AUG. A schematic of the latter alternative form was shown earlier. The location of AAMP at 2q35 places it in a region near the end region of the chromosomal arm where breakpoints are more likely compared to the centromeric region. Importantly, in the sequences of the consensus and variant forms, in-frame codons are present for a heparin binding region that can be translated if an alternative initiating methionine is introduced upstream.

References


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